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BOTANICAL ANTHELMINTICS***Dr. Veerakumari, L.**

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19th August, 2015Accepted 30th September, 2015Published online 17th October, 2015**Key words:**Botanical anthelmintics,
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Helminth parasitism especially gastrointestinal parasitism is one of the major health problems severely limiting the animal productivity in dairy animals. Chemotherapy is a major treatment modality used for the control of helminth infection in livestock. However, due to increasing development of anthelmintic resistance and the limited availability of commercial drugs to the rural people as well as the high cost of such synthetic medicines, a growing interest in the ethno-veterinary approach to examine the anthelmintic properties of plants traditionally used by local farmers in different parts of the globe is emerging. Medicinal plants are resources of new drugs and have served through ages, as a constant source of medicaments for the treatment of a variety of diseases and are known to provide a rich source of botanical anthelmintics. A large number of plant products are being used to combat gastro-intestinal parasites of livestock and also humans. Phytotherapeutic drugs are safe, non-toxic, biodegradable and do not leave residues in animals products. This review paper summarizes the anthelmintic activities of the medicinal plants and possible mode of action which could be further explored for the identification of lead compounds and development of potential anthelmintic drug.

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INTRODUCTION

Livestock, an adjunct to agriculture, plays an important role in the future growth and development of Indian economy. India has the largest livestock population which constitutes nearly 7% towards its national income where cattle and buffaloes generate 54% of energy for agricultural operations. The livestock provides regular employment to 18.4 million people. Helminth parasitism especially gastrointestinal parasitism is one of the major health problems severely limiting the animal productivity in dairy animals. Any effort to intensify livestock production has been seriously affected by various factors, of which the most serious losses are due to gastrointestinal (GI) helminths.

Paramphistomosis and haemonchosis are the major parasitic diseases reducing the production potential of livestock. Clinical paramphistomosis is associated with few species of flukes namely *Cotylophoron corylophorum*, *Fischoederius elongatus*, *Gastrophylax crumenifer*, *Gigantocotyle explanatum*, *Paramphistomum cervi* and *Paramphistomum epilectum*. Clinical signs include anorexia, weight loss, foetid diarrhea, dehydration and death. Pathogenesis due to *Haemonchus contortus* is associated with the haematophagous activities of developing larvae and adult worms. The greatest losses associated with helminth infection are sub-clinical, and economic assessments show that financial costs of internal

parasitism are enormous due to increase in mortality, and a reduction in growth rate and wool production. Chemotherapy is a major treatment modality used for the control of helminth infection in livestock. However, chemotherapeutic anthelmintic drugs for the control of helminth infection are far from satisfaction, probably due to the absence of broad-spectrum activity of these drugs. Resistance to chemotherapeutic anthelmintics is an increasing problem in many countries (Waller and Prichard, 1985). Furthermore, side effects on the host are a serious problem with many chemotherapeutic anthelmintics currently available. It is imperative to decrease the reliance on these chemotherapeutic drugs for parasite control, not only because of resistance, but also because of growing concerns about the adverse consequences of these anti parasitic drugs on the ecosystem and biodiversity. This danger has given impetus to the search for new drugs, with attention focusing on the search for plant products and the application of plant products as alternative methods of control. Environmental aspects favouring plant anthelmintics include their biodegradability.

On the other hand, chemotherapeutic anthelmintics poses the problem of soil contamination and also residual amounts of these anthelmintics may be found in the faeces after treatment and these may have deleterious effects on the environment. Naturally produced plant anthelmintics offer an alternative that can overcome some of these problems and is both sustainable and environmentally acceptable. In response to the need to discover new anthelmintic drugs of natural origin with possibly low toxicity to host animals and also to raise them

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free from chemical inputs, there is a growing interest in ethno medical and ethno veterinary practices across the world (Guarrera, 1999). In this regard, several medicinal plants have been investigated for their anthelmintic properties. However, scientific evidence on the anthelmintic efficacy of most plant products is limited, regardless of their wide ethno veterinary usage. Scientific validation of anthelmintic effects and possible side-effects of plant products in ruminants is necessary prior to their adoption as a novel method for parasite control.

Validation of anthelmintic activities of plants

Pharmacodynamic interaction of drug or plant extracts can be better described using both *in vitro* and *in vivo* studies. *In vitro* studies offer rapid, sensitive and more economical methods for screening and evaluation of anthelmintic activity of plant extracts. Most of the effects recorded *in vitro* also occur *in vivo* (Rahman and Bryant, 1977) and contribute to the anthelmintic efficacy.

Table 1. List of plants with anthelmintic activity

Plant	Part Used	Target parasite	Host	Reference
<i>Acacia albida</i>	Seeds	Worm infestation	Sheep, goat	Nwude and Ibrahim(1980)
<i>Acacia arabica</i> willd.	Root, stem, bark	G.I nematodes	Sheep	Tariq and Tantry (2012)
<i>Acacia Arabica</i>	Bark	<i>Cotylophoron cotylophorum</i>	Sheep	Veerakumari <i>et al.</i> (2012a)
<i>Acacia Arabica</i>	Bark	<i>C. cotylophorum</i>	Sheep	Veerakumari <i>et al.</i> (2013)
<i>Acacia concinna</i>	Pods	<i>C. cotylophorum</i>	Sheep	Priya <i>et al.</i> (2013)
<i>Acacia karoo</i>	Ethanol extract	<i>H. contortus</i>	Goat	Kahiya <i>et al.</i> (2003)
<i>Acacia mearnsii</i>	Leaves extract	<i>H. contortus, Oesophagostomum, Trichostrongylus and Cooperia</i>	Sheep and goat	Max <i>et al.</i> (2007)
<i>Acacia polyacantha</i>	Leaves extract	<i>H. contortus, Oesophagostomum, Trichostrongylus and Cooperia</i>	Sheep and goat	Max <i>et al.</i> (2007)
<i>Acacia tortilis</i>	Leaves extract	<i>H. contortus, Oesophagostomum, Trichostrongylus and Cooperia</i>	Sheep and goat	Max <i>et al.</i> (2007)
<i>Achillea millifolium</i> Linn.	Whole herb	G.I nematodes	Sheep	Tariq and Tantry (2012)
<i>Aconitum chasmantum</i> Stapf.	Root	G.I nematodes	Sheep	Tariq and Tantry (2012)
<i>Aconitum heterophyllum</i> Wall. ex. Royle.	Root	G.I nematodes	Sheep	Tariq and Tantry (2012)
<i>Aconitum violaceum</i>	Root	G.I nematodes	Sheep	Tariq and Tantry (2012)
<i>Acorus calamus</i> Linn.	Rhizome	G.I nematodes	Sheep	Tariq and Tantry (2012)
<i>Adhatoda vasica</i>	Roots	Mixed GI G.I nematodes	Sheep	Lateef <i>et al.</i> (2003)
<i>Adhatoda vasica</i>	Leaves	<i>C. cotylophorum</i>	Sheep	Jeya and Veerakumari (2015)
<i>Ajuga bracteosa</i> Wall.	Whole herb	G.I nematodes	Sheep	Tariq and Tantry (2012)
<i>Albizia anthelmintica</i>	Bark and root	<i>Haemonchus contortus</i> and <i>Trichuris ovis</i>	Cattle, goat, sheep	Minja (1989); ITDG and IIRR (1996)
		<i>Fasciola</i> , Lungworms		
<i>Albizia anthelmintica</i>	Bark	Anthelmintic	Cattle, goat, sheep	Minja (1989); ITDG and IIRR, (1996)
<i>Albizia anthelmintica</i>	Bark	<i>Fasciola gigantica</i>	Goat	Koko <i>et al.</i> (2000)
<i>Albizia coriavera</i>	Bark	<i>Fasciola</i> spp., Lungworms	Cattle, goat, sheep	ITDG and IIRR (1996)
<i>Albizia lebbek</i>	Leaves extract	<i>H. contortus, Oesophagostomum, Trichostrongylus and Cooperia</i>	Sheep and goat	Max <i>et al.</i> (2007)
<i>Alstonia boonei</i>	Bark	<i>Ascaris suum</i>	Cattle, goat, sheep	Fakae <i>et al.</i> (2000)
<i>A. sativum</i>	Bulb	<i>Onchocerus volvulus</i>	Cattle, goat, sheep	ITDG and IIRR (1996)
<i>A. sativum</i>	Bulb	Round-worms	Cattle, goat, sheep	Iqbal <i>et al.</i> (2001)
<i>A. sativum</i>	Bulb/alcoholic extract	<i>Fasciola gigantica, Gigantocotyle explanatum</i>	Cattle, goat, sheep	Singh <i>et al.</i> (2007)
<i>A. sativum</i>	Bulbs	GI nematodes	Goat	Amin <i>et al.</i> (2009)
<i>A. sativum</i>	Bulb/aqueous extract	<i>Fasciola gigantica</i>	Cattle	Jeyathilakan <i>et al.</i> (2012)
<i>A. sativum</i>	Bulb/Aqueous extract	<i>Bunostomum trigonocephalum</i>	Cattle, goat, sheep	Khobragade <i>et al.</i> (1994)
<i>A. sativum</i>	Bulb/Oil extract	<i>Asaridia galli, Heterakis gallinarum</i>	Cattle, goat, sheep	Singh and Nagaich (1999)
<i>A. sativum</i>	Bulb/Methanol extract	<i>H. contortus</i>	Goat, sheep	Iqbal <i>et al.</i> (2001)
<i>A. sativum</i>	Bulbils/ Aqueous extract	<i>H. contortus</i>	Sheep	Veerakumari and Lakshmi (2006)
<i>A. sativum</i>	Bulbils/Aqueous extract	<i>C. cotylophorum</i>	Sheep	Veerakumari <i>et al.</i> (2012b)
<i>A. sativum</i>	Bulb	G.I nematodes	Sheep	Tariq and Tantry (2012)
<i>Amaranthus spinosus</i>	Ethanol extract	GI nematodes	Goat	Sujon <i>et al.</i> (2008)
<i>Ananas comosus</i>	Leaves extract	<i>Toxocara vitulorum</i>	Calves	Islam <i>et al.</i> (2005)
<i>A. comosus</i>	Ethanol extract	GI nematodes	Goat	Sujon <i>et al.</i> (2008)
<i>A. comosus</i>	Leaves	<i>Trichuris ovis and Trichuris globosa</i>	Ruminants	Behnke <i>et al.</i> (2008)

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<i>A. comosus</i>	Leaves	GI nematodes	Goat	Amin et al. (2009)
<i>Andrographis paniculata</i>	Leaves/Aqueous extract	<i>H. contortus</i>	Sheep and goat	Singh et al. (2011)
<i>Areca catechu</i>	Nut	Taenicidal	Cattle, goat, dog	Roepke (1996)
<i>Areca caetcheu</i>	Nut, Ethanol extract	<i>C. cotoylophorum</i>	sheep	Manoj Dhanraj and Veerakumari (2015 a)
<i>Artemesia absinthium</i>	Root powder/aqueous extract	G.I nematodes	Sheep	Tariq et al. (2009)
<i>A. maritime</i>	Whole herb	<i>Neoascaris vitulorum</i>	Buffalo, calves	Akhtar et al. (1982); Farnsworth et al. (1985); Sherif et al. (1987)
<i>A. maritime</i>	Whole herb	G.I nematodes	Sheep	Tariq and Tantry (2012)
<i>A. absinthium</i>	-	G.I nematodes	Sheep	Tariq and Tantry (2012)
<i>A. brevifolia</i>	Crude methonolic and aqueous extract of whole plant	<i>H. contortus</i>	Sheep	Iqbal et al. (2004)
<i>A. herbaalba</i>	Powdered shoots	<i>H. contortus</i>	Sheep and goat	Idris et al. (1982)
<i>A. maritime</i>	Whole herb	<i>Neoascaris vitulorum</i>	Buffalo, calves	Akhtar et al. (1982); Farnsworth et al. (1985); Sherif et al. (1987)
<i>A. maritime</i>	Root powder	<i>H. contortus</i>	Sheep and goat	Jangde et al. (2001)
<i>Artemisia mesatlantica</i>	Flavonoids and sesquiterpene lactones	Anthelmintic	Sheep and goat	Holeman et al. (1991)
<i>Asparagus filicinus</i> Buch.	Root	G.I nematodes	Sheep	Tariq and Tantry (2012)
<i>Azadirachta indica</i>	Fruit seed and oil	G.I nematodes (<i>Trichostongylids</i>)	Small ruminants	Mostofa et al. (1996)
<i>A. indica</i>	Leaves/methanol extract	<i>H. contortus</i>	Goat	Arora et al. (2010)
<i>A. indica</i>	Cake and Leaves	Anthelmintic	Small ruminants	Gowda (1997); Mostofa et al. (1996)
<i>A. indica</i>	Leaves/Ethanol extract	<i>Trichuris globulosa</i>		Sarwal et al. (2000)
<i>A. indica</i>	Leaves extract	<i>H. contortus, Oesophagostomum, Trichostrongylus</i> and <i>Cooperia</i>	Sheep and goat	Max et al. (2007)
<i>A. indica</i>	Bark/Alcoholic extract	<i>H. contortus</i>	Sheep	Swarnkar et al. (2008a)
<i>A. indica</i>	Leaves/aqueous extract	<i>H. contortus</i>	Sheep and goat	Singh et al. (2008)
<i>A. indica</i>	Metanol extract	<i>Fasciola gigantica</i>	Livestock	Kushwaha et al. (2004)
<i>A. indica</i>	Ethanol extract	<i>C. cotoylophorum</i>	Sheep	Veerakumari and Priya (2006)
<i>A. indica</i>	Leaves	GI nematodes	Goat	Amin et al. (2009)
<i>Balanites aegyptiaca</i>		<i>Fasciola gigantica</i>	Goat	Koko et al. (2000)
<i>Bauhinia tomentosa</i>	Leaves/Aqueous and ethanol extracts	<i>H. contortus</i>	Sheep and goat	Vihan et al. (2007)
<i>Bauhinia variegata</i>	Leaves/Methanol extract	<i>H. contortus</i>	Sheep and goat	Bharadwaj et al. (2010)
<i>Boswellia dalzielii</i>	Bark	Anthelmintic	Sheep, goat	Nwude and Ibrahim (1980)
<i>Brassica nigra</i> Linn.	Seed	G.I nematodes	Sheep	Tariq and Tantry (2012)
<i>Butea frondosa</i>	Seeds	<i>Ascaridia galli</i>	In vitro	Lal et al. (1976)
<i>B. frondosa</i>	Seed/ether extract	<i>Ascaridia galli</i>		Sharma and Sisodia (1976)
<i>B. frondosa</i>	Seeds/Palasonin	<i>Ascaris lumbicooides</i>	Sheep	Rao et al. (1977)
<i>B. frondosa</i>	Seeds/Isolated a purified/Palasonin	<i>Ascaridia galli</i>	Livestock	Kumar et al. (1983)
<i>B. frondosa</i>	Seeds/Solution in ammonia (Palasonin)	<i>H. contortus</i>	Sheep	Sathianesan et al. (1984)
<i>B. frondosa</i>	Seeds/Ether and Alcoholic extracts	<i>Ascaridia galli</i>	Livestock	Shilaskar and Parashar (1989)
<i>B. frondosa</i>	Seeds/Aqueousextract (Palasonin)	<i>H. contortus</i>	Sheep	Jangde et al. (2001)
<i>B. frondosa</i>	Seeds/Aqueous, alcohol, ether and chloroform	<i>H. contortus</i>	Sheep	Swarnkar et al. (2008c)
<i>B. frondosa</i>	Seeds/Chloroform extract	<i>H. contortus</i>	Sheep	Vihan et al. (2007)
<i>B. frondosa</i>	Seeds/Palasonin	<i>Ascaris lumbicooides</i>	Sheep	Parker and Palmer (1991)

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<i>B. monosperma</i>	Seeds	Anthelmintic, GI G.I nematodes	Sheep and others	Kalesaraj and Kurup(1968); Chandra and Sabir(1978); Lal et al.(1978); Prashanth et al. (2001) Iqbal et al. (2006)
<i>B. monosperma</i>	Seeds	<i>Paramphistomum cervi</i>	Goat	Chopra et al.(1991)
<i>Caesalpinia crista</i>	Seed powder	<i>Fasciola gigantica</i>	Buffalo	Kailani et al. (1995)
<i>Caesalpina crista</i>	Seeds	<i>Toxocara vitulorum</i>	Buffalo, calves, chicken	Akhtar et al. (1985); Javed et al. (1994)
<i>Ascaridia galli</i>				
<i>Calliandra portoricensis</i>	Roots, leaves, flowers	<i>Toxocaracanis</i> , Gastrointestinal G.I nematodes, <i>H. contortus</i>	Dog, Sheep	Adeunumi and Akubue (1981); Jain et al.(1996); Al- Qarawi et al. (2001); Iqbal et al. (2005)
<i>Calotropis procera</i>	Latex	Gastrointestinal nematodes, <i>H. contortus</i>	Sheep	Iqbal et al. (2005)
<i>Calatropis gigantean</i>	Leaves/Extract (Methanol)	<i>H. contortus</i>	Sheep and goat	Yadav et al. (2010)
<i>Cannabis sativa</i> Linn.	Leaf	G.I nematodes	Sheep	Tariq and Tantry (2012)
<i>Capillipedium foetidum</i>	Oil, grass	<i>Taenia</i> <i>solum</i> and <i>Ascaris</i> <i>lumbricooides</i> <i>Ascaridia galli</i>	<i>In vitro</i>	Siddiqui and Garg (1990)
<i>Carica papaya</i>	Seeds/Alcoholic extract		Small ruminants	Lal et al. (1976)
<i>C. papaya</i>	Seeds/Aqueous extract	<i>A.galli</i> and <i>Heterakis gallinae</i>	Small ruminants	Singh and Nagaich (1999)
<i>C. papaya</i>	Leaves	<i>Trichuris ovis</i> and <i>Trichuris</i> <i>globosa</i>	Ruminants	Behnke et al. (2008)
<i>C. papaya</i>	Seeds	GI nematodes	Goat	Amin et al. (2009)
<i>C. papaya</i>	Leaves	GI nematodes	Goat	Amin et al. (2009)
<i>Carissa edulis</i>	Root	Roundworms	Cattle, goat, sheep	ITDG and IIRR (1996)
<i>Casia tora</i>	Seeds/Alcohol and aqueous extracts	<i>Ascaridia galli</i>	Small ruminants	Deore et al. (2009)
<i>Cassia spectalis</i>	Roots	Round-worms	Cattle, goat,	ITDG and IIRR (1996)
<i>Chenopodium album</i>	Leaves	G.I nematodes	Sheep	Akhtar et al. (1999)
<i>Chenopodium album</i>	Seeds/Methanol extract	<i>H. contortus</i>	Sheep	Yadav et al. (2010)
<i>Chenopodium ambrosoides</i>	Whole herb	G.I nematodes	Sheep	Tariq and Tantry (2012)
<i>Linn.</i>				
<i>Chrysophyllum cainito</i>	Stem	<i>H. contortus</i>	Cattle	Fernandez (1991)
<i>Chrysanthmnus viscidiflorus</i>	Methanol extract	<i>H. contortus</i>	Goat, sheep	Acharya et al. (2014)
<i>Cissampelos mucromata</i>	Root	Helminths	Goat, sheep	Minja (1989)
<i>Cimum basilicum</i>	Leaves	GI nematodes	Goat	Amin et al. (2009)
<i>Citrus aurantifolia</i>	Leaves	GI nematodes	Goat	Amin et al. (2009)
<i>Corchorus spp.</i>	Ethanol extract	GI nematodes	Goat	Sujon et al. (2008)
<i>Corchorus capsularis</i>	Leaves	GI nematodes	Goat	Amin et al. (2009)
<i>Croton macrostachys</i>	Leaves	Anthelmintic	Goat	Minja (1989)
<i>Curcuma longa</i>	Rhizome	GI nematodes	Goat	Amin et al. (2009)
<i>Cucurbita Mexicana</i>	Methanol extract	<i>H. contortus</i>	Sheep and goat	Iqbal et al. (2001)
<i>Cucurbita sp.</i>	Methanol extract	G.I nematodes	Sheep	Tariq and Tantry (2012)
<i>Cuscuta reflexa</i>	Whole plant	GI nematodes	Goat	Amin et al. (2009)
<i>Cyathoclina lyrata</i>	Essential oil	Tape-worms, hookworms	<i>In vitro</i>	Shrivastava (1979)
<i>Cymbopogon flexuosus</i>	Leaves/Essential Oil	Strongyles	Goat	Sahoo and Tripathy (2007)
<i>Cynodon dactylon</i>	Ethanol extract	G.I nematodes	Goat, sheep	Sujon et al. (2008)
<i>Datura quercifolia</i>	Fruit	<i>Ascaridia galli</i>	<i>In vitro</i>	Kaushik et al., 1981
<i>Datura stramonium</i> Linn.	Seeds	G.I nematodes	Sheep	Tariq and Tantry (2012)
<i>Daucus carota</i> Linn.	Roots	G.I nematodes	Sheep	Tariq and Tantry (2012)
<i>Delphinium denudatum</i> Wall.	Seeds	G.I nematodes	Sheep	Tariq and Tantry (2012)
<i>Diospyros mespiliiformis</i>	Bark	Anthelmintic	Sheep, goat	Minja (1989)
<i>Diospyros scabra</i>	Seeds	<i>Fasciola</i>	Cattle, goat, sheep	ITDG and IIRR (1996)
<i>Dodonea viscosa</i>	Leaves	Intestinal worms	Cattle	Sharma and Singh (1989)
<i>Embelia kilimandschiraca</i>	Root	Helminthosis	Cattle	Minja (1989)
<i>Embelia ribes</i>	Seeds/Aqueous extract	<i>H. contortus</i>	Sheep and goat	Swarankar et al. (2009)
<i>E. ribes</i>	Aqueous extract	<i>H. contortus</i>	Goat	Arora et al. (2010)
<i>E. ribes</i>	Seed/Aqueous and Alcoholic extracts	<i>Paramphistomum cervi</i>	Goat, sheep	Chopra et al.(1991)
<i>E. ribes</i>	Seed/Aqueous and Alcoholic extracts	<i>H. contortus</i>	Sheep	Swarankar et al. (2009)
<i>Ericameria nauseosa</i>	Methanol extract	<i>H. contortus</i>	Goat, sheep	Acharya et al. (2014)
<i>Erythrina senegalensis</i>	Bark	<i>Fasciola</i> spp.	Ruminants	Nwude and Ibrahim (1980)
<i>Eugeniu</i> spp.	Ethanol extract	GI nematodes	Goat	Sujon et al. (2008)
<i>Ferula asafetida</i>	Extract	<i>Ascaridia galli</i> and <i>Heterakis gallinae</i>	Livestock	Nagaich and Singh (2001)

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<i>Ferula costata</i>	Crude methanol and aqueous extract	<i>H. contortus</i>	Sheep	Kakar <i>et al.</i> (2013)
<i>Ficus religiosa</i>	Leaves/Methanol extract	<i>H. contortus</i>	Goat, sheep	Iqbal <i>et al.</i> (2001)
<i>Ficus spp</i>	Leaves	<i>Trichuris ovis</i> and <i>Trichuris globosa</i>	Ruminants	Behnke <i>et al.</i> (2008)
<i>Flemingia vestita</i>	Tuber/Extract (genistein)	<i>Ascaris suum</i>	Small ruminants	Yadav <i>et al.</i> (1992)
<i>Flemingia macrophylla</i>	Leaves	<i>H. contortus</i>	Sheep	Nyguyen <i>et al.</i> (2005)
<i>Fritillaria roylei</i> Hook.	Bulb	G.I nematodes	Sheep	Tariq and Tantry (2012)
<i>Fumaria parviflora</i>	Plant	<i>Tricho strongylus</i> , <i>Haemonchus</i> ,	Sheep, buffalo	Akhtar and Javed, (1985); Kailani <i>et al.</i> (1995)
	Powder	<i>Trichuris, Fasciola</i> spp.		
<i>Gardenia gummiifera</i>		Round worm	Goats	Jangde and Bansod (2004)
<i>Geranium viscosissimum</i>	Methanol extract	<i>H. contortus</i>	Goat, sheep	Acharya <i>et al.</i> (2014)
<i>Gliricidia sepium</i>	Leaves extract	<i>H. contortus</i> , <i>Oesophagostomum</i> , <i>Trichostrongylus</i> and <i>Cooperia</i>	Sheep and goat	Max <i>et al.</i> (2007)
<i>Gmelina arborea</i>	Leaves	GI nematodes	Goat	Amin <i>et al.</i> (2009)
<i>Hagenia abyssinica</i>	Fruit	Round-worms	Cattle, goat, sheep	ITDG and IIRR, 1996
<i>Holarrhena antidysenterica</i>	Leaves	GI nematodes	Goat	Amin <i>et al.</i> (2009)
<i>Hyoscyamus niger</i>	Seeds	Mixed nematode Infection	<i>In vivo</i>	Akhtar and Ahmad, 1990
<i>Inula racemosa</i> Hook.	Root	G.I nematodes	Sheep	Tariq and Tantry (2012)
<i>Istonia scholaris</i>	Leaves	G.I nematodes	Goat	Amin <i>et al.</i> (2009)
<i>Juglans regia</i> Linn.	Bark and Leaf	G.I nematodes	Sheep	Tariq and Tantry (2012)
<i>Juniperus macropoda</i> Boiss.	Leaf	G.I nematodes	Sheep	Tariq and Tantry (2012)
<i>Juniperus communis</i> Linn.	Fruit	G.I nematodes	Sheep	Tariq and Tantry (2012)
<i>Khaya senegalensis</i>	Bark	<i>H. contortus</i> <i>Trichostrongylus</i> Sp. <i>Oesophagostomum</i> Sp. <i>Strongyloides</i> Sp. <i>Trichuris</i> Sp.	Sheep	Ademola <i>et al.</i> (2004)
<i>Lagen siceraria</i>	Seeds	Cestodes, <i>moniezia</i> , <i>avitelina</i> spp.	Sheep	Akhtar and Riffat, 1987
<i>Lansium domesticum</i>	Seeds	<i>Ascaridia galli</i> <i>Ascaris suum</i>	Goat	Fernandez (1991)
<i>Lantana trifolia</i>	Fruit	<i>H. contortus</i> <i>Fasciola</i> spp., lungworms	Cattle, goat, sheep	ITDG and IIRR, 1996
<i>Lawsonia inermis</i>	Leaves	<i>Fasciola</i> spp.	Sheep, goat	Nwude and Ibrahim, 1980
<i>Lawsonia inermis</i>	Leaves	<i>Setaria digitata</i>	-	Banu <i>et al.</i> (1992)
<i>Leucas aspera</i>	Leaves	GI nematodes	Goat	Amin <i>et al.</i> (2009)
Leucena leucocephala and	Leaves	Mixed GI infection	Small	Semenye, 1990; Adejumo and Ademosun, 1991).
Leucena diversifolia			ruminants	Fernandez (1991)
<i>Leucaena leucocephala</i>	Seeds	<i>Ascaridia galli</i> <i>Ascaris suum</i>	Chicken	
		<i>H. contortus</i>	Pig,	
			Goat	
<i>Lespedeza cuneata</i>	fresh forage or hay	<i>H. contortus</i>	Sheep	(Min <i>et al.</i> , 2004; Lange <i>et al.</i> , 2006; and Shaik <i>et al.</i> , 2006)
<i>Liatris punctate</i>	Methanol extract	<i>H. contortus</i>	Goat, sheep	Acharya <i>et al.</i> (2014)
<i>L. corniculatus</i>	Methanol extract	<i>H. contortus</i>	Goat, sheep	Acharya <i>et al.</i> (2014)
<i>Lotus pedunculatus</i>		<i>Teladorsagia circumcincta</i>	Sheep	Niezen <i>et al.</i> (1998)
<i>Luvunga scandens</i>	Ethanol extract	<i>H. contortus</i>		Sujon <i>et al.</i> (2008)
<i>Mallotus philippinensis</i>	Leaves/Chloroform extract	<i>H. contortus</i>	Sheep and goat	Vihan <i>et al.</i> (2007)
<i>M. philippinensis</i>	Fruit	<i>Moniezia</i> and <i>Avitellina</i>	Beetal Goat	Akhtar and Ahmad (1992)
<i>M. philippinensis</i>	Essential oil	<i>Fasciola gigantica</i>	Sheep and goat	Kushwaha <i>et al.</i> (2004)
<i>Momordica charantias</i>				

Continue

<i>Mangifera indica</i>	Seeds	<i>Ascaridia galli</i>	Chicken	Fernandez (1991)
<i>Magnifera indica</i>	Ethanol extract	G.I nematodes	Goat	Sujon <i>et al.</i> (2008)
<i>Manihot esculenta</i>	Leaves	<i>H. contortus</i>	Sheep	Nyguyen <i>et al.</i> (2005)
<i>Matricaria chamomilla Linn.</i>	Aerial parts	G.I nematodes	Sheep	Tariq and Tantry (2012)
<i>Melia azedarach</i>	Fruit (Powdered)	GI. nematodes	Goat	Akhtar and Riffat (1984)
<i>M. azederach</i>	Fruit	<i>Ascaridia galli</i> <i>Haemonchus</i> , <i>Trichostrongylus</i> , <i>Trichuris</i> , <i>Chabertia spp.</i>	Goat	Akhtar and Riffat (1985);
<i>M. azedarach</i>	Seed/Alcoholic extract	<i>H. contortus</i>	Goat	Misri <i>et al.</i> (2002)
<i>M. azedarach</i>	Bark	GI nematodes	Goat	Amin <i>et al.</i> (2009)
<i>M. azedarach</i>	Leaves	GI nematodes	Goat	Amin <i>et al.</i> (2009)
<i>Melilotus alba</i>	Methanol extract	<i>H. contortus</i>	Goat, sheep	Acharya <i>et al.</i> (2014)
<i>Melilotus officinalis</i>	Methanol extract	<i>H. contortus</i>	Goat, sheep	Acharya <i>et al.</i> (2014)
<i>Mentha sylvestris Linn.</i>	Whole herb	G.I nematodes	Sheep	Tariq and Tantry (2012)
<i>Mimisa isp</i>	Leaves	<i>H. contortus</i>	Sheep	Nyguyen <i>et al.</i> , 2005
<i>Mimosa pudica</i>	Stem	<i>H. contortus</i>	Sheep and goat	Fernandez (1991)
<i>M. pudica</i>	Leaves	<i>H. contortus</i>	Sheep	Nyguyen <i>et al.</i> (2005)
<i>Momordica charantia</i>	Ethanol extract	GI nematodes	Goat	Sujon <i>et al.</i> (2008)
<i>M. charantia</i>	Leaves	GI nematodes	Goat	Amin <i>et al.</i> (2009)
<i>M. charantia</i>	Stem	<i>Ascaris suum</i> <i>Ascaridia galli</i> <i>H. contortus</i>	Pig, Chicken, Goat	Fernandez (1991); Farnsworth <i>et al.</i> (1985)
<i>Moringa oleifera</i>	Seeds	<i>Ascaridia galli</i> <i>Ascaris suum</i> <i>H. contortus</i>	Chicken, Pig, Goat	Fernandez (1991)
<i>Morus alba</i>	Leaves extract	<i>H. contortus</i> , <i>Oesophagostomum</i> , <i>Trichostrongylus</i> and <i>Cooperia</i>	Sheep and goat	Max <i>et al.</i> , 2007
<i>Morus alba Linn.</i>	Bark and Leaf	Nematodes	Sheep	Tariq and Tantry (2012)
<i>Mixture of</i>	Ethanol extract	<i>H. contortus</i>	Cow claves	Raje <i>et al.</i> (2003)
<i>Azadirachta indica</i>		<i>Paramphistomum cervi</i>		
<i>Butea frondosa</i>		<i>Oesophagostomum</i>		
<i>Nigella sativa</i>				
<i>Piper longum</i>				
<i>Myrsine africana Linn.</i>	Whole plant	Nematodes	Sheep	Tariq and Tantry (2012)
<i>Myrsine africana</i>	Leaf	Roundworms	Cattle, goat, sheep,	ITDG and IIRR (1996)
<i>Nauclea latifolia</i>	Roots	<i>Ascaris suum</i> <i>Onchocerous volvulus</i>	In vitro	Fakae <i>et al.</i> (2000)
<i>Nelumbo nucifera Gaertn.</i>	Whole plant	Nematodes	Sheep	Tariq and Tantry (2012)
<i>Nepeta cataria Linn.</i>	Leaf/whole plants	Nematodes	Sheep	Tariq and Tantry (2012)
<i>Nicotiana tabacum</i>	Dried leaves/ Decoction	<i>H. contortus</i>	Sheep and goat	Raje and Jangde (2003)
<i>Nicotiana tobacum</i>	Dried leaves/ Aqueous and methanol extracts	<i>H. contortus</i>	Sheep and goat	Iqbal <i>et al.</i> (2006)
<i>Nicotiana tobacum</i>	Ethanol extract	GI nematodes	Goat	Sujon <i>et al.</i> (2008)
<i>N. tabacum</i>	Leaves	GI nematodes	Goat	Amin <i>et al.</i> (2009)
<i>Nigella sativa</i>	Seed powder	<i>F. gigantica</i>	Buffalo	Kailani <i>et al.</i> (1995)
<i>Nymphaea alba Linn.</i>	Whole plant	Nematodes	Sheep	Tariq and Tantry (2012)
<i>Ocimum basilicum Linn.</i>	Seeds	Nematodes	Sheep	Tariq and Tantry (2012)
<i>O. gratissimum</i>	Leaves	<i>Ascaris suum</i> <i>Onchocerous volvulus</i>	In vitro	Fakae <i>et al.</i> (2000)
<i>O. sanctum</i>	Leaves	<i>H. contortus</i>	Sheep and goat	Garg (1997)
<i>O. sanctum</i>	Leaves	<i>Setaria digitata</i>	Livestock	Banu <i>et al.</i> (1992)
<i>Onobrychis viciifolia</i>	Hay	GI infections	Goats	Paolini <i>et al.</i> (2004, 2005).
<i>Peganum harmala</i>	Seeds	Mixed GI infection, cestode infection	Goats	Akhtar and Ahmed (1991)
<i>P. harmala</i>	Seeds	Nematodes	Sheep	Tariq and Tantry (2012)
<i>P. harmala</i>	Seeds	Mixed gastrointestinal infection	Goat	Akhtar and Ahmad (1991);
<i>Perideridia gairdneri</i>	Methanol extract	<i>H. contortus</i>	Sheep	Akhtar and Riffat (1986)
<i>Persea americana</i>	Leaves extract	<i>H. contortus</i> , <i>Oesophagostomum</i> , <i>Trichostrongylus</i> and <i>Cooperia</i>	Goat, sheep	Acharya <i>et al.</i> (2014)
<i>Pinus wallichiana</i>	Needles	Nematodes	Sheep	Tariq and Tantry (2012)
<i>Piper betle</i>	Leaves/Essential oil	<i>Ascaridia galli</i>	Sheep	Shilaskar and Parashar (1989)
<i>P. betle</i>	Ethanol extract	GI nematodes	Goat	Sujon <i>et al.</i> (2008)
<i>P. betle</i>	Leaves	GI nematodes	Goat	Amin <i>et al.</i> (2009)

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<i>Piliostigma thonningii</i>	Bark, leaves, Roots and bark extract	<i>Ascaris suum</i> <i>Onchocerus volvulus</i>	Ruminants	Fakae <i>et al.</i> (2000)
<i>Ocimum gratissimum</i>				
<i>Nauclea latifolia</i>				
<i>Alstonia boonei</i>				
<i>Platanus orientalis</i>	Bark, Seed	Nematodes	Sheep	Tariq and Tantry (2012)
<i>Pongamia glabra</i>	Seeds/Ether extract	<i>H. contortus</i>	Sheep and goat	Arora <i>et al.</i> (2010)
<i>Populus alba</i> Linn.	Bark	Nematodes	Sheep	Tariq and Tantry (2012)
<i>Populus nigra</i> Linn.	Bark	Nematodes	Sheep	Tariq and Tantry (2012)
<i>Prosopis cineraria</i>	Bark	<i>H. contortus</i>	Sheep and goat	Swarnker <i>et al.</i> , 2008(b)
<i>P. cineraria</i>	Bark, etanol extract	<i>C. corylophorum</i>	Sheep	Manigandan and Veerakumari (2015)
<i>Prunella vulgaris</i>	Root and stem bark	Nematodes	Sheep	Tariq and Tantry (2012)
<i>Psidium guajava</i>	Leaves	<i>H. contortus</i>	Sheep	Nyguyen <i>et al.</i> (2005)
<i>P. guajava</i>	Leaves extract	<i>H. contortus, Oesophagostomum, Trichostrongylus and Cooperia</i>	Sheep and goat	Max <i>et al.</i> (2007)
<i>Punica granatum</i>	Leaves	GI nematodes	Goat	Amin <i>et al.</i> (2009)
<i>P. granatum</i>	Rind	<i>C. corylophorum</i>	Sheep	Veerakumari <i>et al.</i> (2012 b)
<i>P. granatum</i> Linn.	Flowers and leaves	G.I nematodes	Sheep	Tariq and Tantry (2012)
<i>Quisqualis indica</i>	Seeds	<i>H. contortus</i>	Goat	Xiao and Lin (1986)
<i>Quisqualis indica</i>	Stem	<i>Ascaris suum</i> <i>Ascaridia galli</i> <i>H. contortus</i>	Pig, Chicken, Goat	Fernandez (1991); Farnsworth <i>et al.</i> (1985)
<i>Raphanus sativus</i>	Root	Nematodes	Sheep	Tariq and Tantry (2012)
<i>Rapanea melanoploeos</i>	Seeds	Roundworms	Cattle, goat, sheep	ITDG and IIRR (1996)
<i>Rhamnus principipes</i>	Leaves	Anthelmintic	Cattle, goat, sheep	Minja (1989)
<i>Rhus vulgaris</i>	Root	Roundworms	Cattle, goat, sheep	ITDG and IIRR (1996)
<i>Rhus vulgaris</i>	Roots	Round-worms	Cattle, goats, sheep	ITDG and IIRR (1996)
<i>Rhus aromatic</i>	Methanol extract	<i>H. contortus</i>	Goat, sheep	Acharya <i>et al.</i> (2014)
<i>Roltleria tinctoria</i>		<i>Paramphistomum cervi</i>	Sheep and goat	Chopra <i>et al.</i> (1991)
<i>Salvia plebeja</i>	Leaves	GI nematodes	Goat	Amin <i>et al.</i> (2009)
<i>Salix alba</i> Linn.	Bark	Nematodes	Sheep	Tariq and Tantry (2012)
<i>Sanguinaria canadensis</i>	Methanol extract	<i>H. contortus</i>	Goat, sheep	Acharya <i>et al.</i> (2014)
<i>Senecio lyratiparitus</i>	Leaves	Anthelmintic	Goat, sheep	Minja (1989)
<i>Schinopsis</i> sp	Bark extract	<i>H. contortus</i>	Goat and sheep	Paolini <i>et al.</i> (2004)
<i>Skimmia laureola</i>	Essential oil from root, stem and leaves	<i>H. contortus</i>	Sheep	Mehmood <i>et al.</i> (2011)
<i>Solanum nodiflorum</i>	Fruit	Worm infestation	Sheep and goat	Nwude and Ibrahim (1980)
<i>Spigelia anthelmia</i>	Aerial parts	<i>H. contortus</i>	Sheep and goat	Assis <i>et al.</i> (2003)
<i>Stephania glabra</i>	Ethanol extract	<i>Fasciolopsis buski</i> <i>Raillietina echinobothrida</i>	Livestock	Tandon <i>et al.</i> (2004)
<i>Syzygium aromaticum</i>	Bud	<i>C. corylophorum</i>	Sheep	Manoj Dhanraj and Veerakumari (2014;2015 a, b)
<i>Tagetes patula</i>	Flowers/Methanol extract	<i>H. contortus</i>	Sheep and goat	Bhardwaj <i>et al.</i> (2010)
<i>T. patula</i>	Flowers/Methanol extract	<i>H. contortus</i>	Sheep and goat	Singh <i>et al.</i> (2002)
<i>Tamarindus indica</i>	Leaves extract	<i>H. contortus, Oesophagostomum, Trichostrongylus and Cooperia</i>	Sheep and goat	Max <i>et al.</i> (2007)
<i>T. indica</i>	Root	Roundworms	Cattle, goat, sheep,	ITDG and IIRR (1996)
<i>Terminalia arjuna</i>	Bark/Crude methanolic extract	<i>H. contortus</i>	Sheep	Bachaya <i>et al.</i> (2009)
<i>Terminalia chebula</i>	Dry fruits	<i>Cotylophoron corylophorum</i>	Sheep	Lokesh and Veerakumari(2015)
<i>Thuja sinensis</i>	Leaves/Ethanol extract	<i>H. contortus</i>	Sheep and goat	Vihan <i>et al.</i> (2007)
<i>Tiunospora rumphii</i>	Stem	<i>H. contortus</i>	Goat	Fernandez (1991)
<i>Trachispermum ammi</i>	Seeds/Extract with various solvents (Ethanol, methanol)	<i>Setaria cervi</i>	Sheep	Rizvi <i>et al.</i> (2012)
<i>Trachyspermum ammi</i>	Seed/Extract	<i>H. contortus</i>	Sheep	Lateef <i>et al.</i> (2006)
<i>Trichilia emetic</i>	Bark	<i>Fasciola spp. lungworms</i>	Cattle , goats,sheep	ITDG and IIRR (1996)

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<i>Trichosanthes multiloba</i>		<i>Fasciolopsis buski</i> <i>R. echinobothrida</i> <i>Gastrothylax crumenifer</i>	Livestock	Tandon <i>et al.</i> (2004)
<i>Trigonella foenum-graecum</i>	Seeds		Buffaloes	Swarnakar <i>et al.</i> (2014)
<i>Urtica dioica</i> Linn. <i>Uvaria hookeri</i>	Leaf Root-bark	Nematodes <i>H. contortus</i>	Sheep Sheep and goat	Tariq and Tantry (2012) Padmaja <i>et al.</i> (1993)
<i>Vernonia amygdalina</i> <i>V. anthelmintica</i>	Stem, bark Seeds/Ether and alcoholic extracts	<i>H. contortus</i> <i>Ascaridia galli</i>	In vitro Livestock	Alawa <i>et al.</i> (2003) Shilaskar and Parashar (1989)
<i>Vitex negundo</i> <i>Xanthium flexuosus</i>	Leaves Leaves/Fresh Juice	GI nematodes <i>H. contortus</i>	Goat Sheep and goat	Amin <i>et al.</i> (2009) Sharma <i>et al.</i> (2003)
<i>Xanthium strumarium</i>		<i>H. contortus</i>	Sheep and goat	Sharma <i>et al.</i> (2003)
<i>Zingiberus officinale</i>	Rhizome/ Methanol extract	<i>H. contortus</i>	Sheep and goat	Iqbal <i>et al.</i> (2001)
<i>Z. officinale</i>	Rhizome/Extract	GI. nematodes	Sheep	Iqbal <i>et al.</i> (2006)

However, concentrations of potentially active substances used *in vitro* do not always correspond to *in vivo* bioavailability. Hence, further *in vivo* study is required to enlighten the therapeutic potential of plant extracts in treating helminth infection. The *in vivo* assay such as faecal egg count reduction test is suitable for the evaluation of all types of anthelmintics (Verma *et al.*, 2006). Further, *in vivo* observations on the haematobiochemical profile can offer more insights into the safety and protective efficacy of the plant extracts.

Many of the *in vitro* investigations on anthelmintic activity of plants, their oils, or extracts have been based on their toxic effects on the earthworm, *Pheritima posthuma* (Gaind and Budhiraja, 1967; Kokate and Varma, 1971; Dixit and Varma, 1975; Banerjee and Nigam, 1978; Girgune *et al.*, 1978; 1979; Garg and Kasera 1982a, b; Nanda *et al.*, 1987; Siddiqui and Garg, 1990; Garg and Siddiqui, 1992). However anthelmintic potential of plants should not be assessed based on their efficacy against earthworm. The anatomy and physiology of earthworm, which is an annelid and terrestrial in habitat differ from helminth parasites. Helminths are endo-parasites protect themselves by the production of anti-enzymes and derive energy through anaerobic metabolism. The plants which are effective against earthworm may not have anthelmintic efficacy. Even among the helminth parasites, the plants which are effective against trematodes are not effective against nematodes or cestodes. Hence using earthworm as a model for testing anthelmintic efficacy is inappropriate. Larval and adult Nematodes, cestodes and trematodes are used for the evaluation of anthelmintic activity of different plant material by various researchers (Aktar *et al.*, 2000; Veerakumari and Priya, 2006; Veerakumari and Lakshmi, 2006).

The most common effect of anthelmintic drugs against worm infections is paralysis of the parasite musculature either by inhibiting the neuromuscular transmission or enzymes involved in energy production; however, certain other drugs damage the tegument allowing partial digestion or rejection by the host immune system (Manger, 1991). The pharmacologic basis of the treatment for helminth infection generally involves interference with the integrity of parasite cells, neuromuscular coordination, energy metabolism or protective mechanisms against host immunity, which lead to starvation, paralysis, and expulsion or digestion of the parasite. A full understanding of the way in which drugs achieve their action helps to identify susceptible parasite systems and show the properties of drugs necessary to attack them.

Usage of plant anthelmintics to combat gastrointestinal helminths of small ruminants is well documented by several parasitologists (Hammond *et al.*, 1997; Schillhorn van Veen, 1997; Akhtar *et al.*, 2000; Tagboto and Towson, 2001; Githori *et al.*, 2006; Piyush *et al.*, 2013; Pathak and Chhabra, 2014). Aqueous and various solvent extracts viz. hexane, chloroform, ethylacetate, alcoholic extracts, pastes, essential oils, extracted from leaves, stem, fruits, flowers and roots or whole plants were used to test their anthelmintic efficacy *in vitro* or *in vivo*. Plants with anthelmintic activity are shown in Table 1.

An anthelmintic drug which could cause adverse changes to a parasite but not the host is said to be potential anthelmintics drug. Anthelmintic drugs impair the vital activities of the parasite and consequently result in the death of the parasites. The most common effect of anthelmintic drug is paralysis of parasite musculature either by inhibiting neuro-muscular transmission or enzymes involved in energy production (Manger, 1991; Veerakumari and Munuswamy, 2000). Drugs may also damage the tegument allowing partial digestion or rejection by host immune system. Anthelmintic drugs may induce pathological changes in various organ systems of the parasites affecting the vital physiological functions viz. excretion, osmoregulation and reproduction (Veerakumari and Munuswamy, 1999; Veerakumari and Paranthaman, 2004).

Mode of action of Phytoconstituents

Whole plant consumption has shown good anthelmintic effects against GI parasites of sheep. This could be either due to direct toxic effects on the parasites or it can be indirect due to changes in the gut environment that favour low fecundity and worm expulsion. The mechanisms whereby the consumption of certain plants and plant extracts can affect parasite viability, motility and fecundity both *in vitro* and *in vivo* are largely unknown. For some plants it has been suggested that the consumption could be associated with an enhanced immune response of host towards the parasites as a result of nutrient supplementation and thus improve nutrition (Athanasiadou and Kyriazakis, 2004). Curative property of the medicinal plants is attributed to the various chemical substances of different composition that arise as secondary metabolites and get localized in one or more parts of the plant. These complex phytochemicals could play a vital role in minimizing the development of drug resistance among parasites. A number of research works have elucidated the anthelmintic effect of tannin rich plants against GI helminth parasites (Athanasiadou,

et al., 2001; Paolini *et al.*, 2003; Hoste *et al.*, 2006; Alonso Díaz *et al.*, 2008; Calderón-Quintal *et al.*, 2010; Martínez-Ortíz-de-Montellano *et al.*, 2010). Apart from condensed tannins, catechins, flavonoids, steroids (Oliveira *et al.*, 2009) and polyphenols (Lorimer *et al.*, 1996) as well as bio-active enzymes such as cysteine proteinase and secondary metabolites such as alkaloids, and glycosides (Piyush Jain *et al.*, 2013) possess anthelmintic property. Phytoconstituents, may act by inhibition of tubulin polymerization and blocking glucose uptake (Jain *et al.*, 2011). Any damage to the mucopolysaccharide membrane of parasites will expose the outer layer restricting their movement which finally may cause paralysis and ultimately death of parasite (Chandrashekhar *et al.*, 2008). The anthelmintic effects of tannins may be attributed to its capacity to bind free protein available for larval nutrition and thus reducing the nutrient availability resulting in larval starvation or decrease in gastrointestinal metabolism directly through inhibition of oxidative phosphorylation causing larval death (Scalbert., 1991; Athanasiadou *et al.*, 2001).

Conclusion

Discovery of new therapeutic substances of natural origin, with possibly least toxicity to animals will protect the animals from chemical inputs. The natural compounds derived from plants are more stable as these are mostly plant secondary metabolites synthesized over a long synthetic ones and therefore are a source of low molecular weight structures active against a wide range of target agents and this diversity can preclude the occurrence of resistance. The progress in the livestock sector will in turn result in more balanced development of rural economy and improvement in economic status of poor people associated with this sector.

A combinatorial approach would help us to formulate new anthelmintic phytomedicine to combat parasitic infections in livestock. Intensive effort on isolation, combined formulation and validation of the efficacy of these plant based anthelmintics and appropriate toxicological screening should be undertaken before the release of commercially viable product for use in livestock industry. Future research in this area seems promising, as there remain a lot of plants to be tested for their anthelmintic potential.

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